An Improved Synthesis of Selectively Protected L-Dopa Derivatives from L-Tyrosine

Chen Chen, Yun-Fei Zhu, and Keith Wilcoxen

Department of Medicinal Chemistry. Neurocrine Biosciences, Inc., 10555 Science Center Drive, San Diego, California 92121

Received August 27, 1999

Many biologically active natural products such as the piperazinomycin (2),¹ K-13 (3a),² OF4949-I-IV (3b-e),³ bouvardin and deoxybouvardin,⁴ and RA-I-IV⁵ contain a key structural unit isodityrosine (1) which is a naturally occurring dimeric form of L-Dopa [L-3-(3,4-dihydroxyphenyl)alanine, 4]. L-Dopa derivatives with two differentiated



protecting groups on the catechol are useful building blocks for synthetic construction of these molecules. Several methods toward the synthesis of selectively protected L-Dopa derivatives have been reported in the literature.⁶⁻⁸ The synthesis reported by Boger and Yohannes⁶ provides the 4-O-benzyl-N-Cbz-L-Dopa methyl

(2) (a) Kase, M.; Kaneko, M.; Yamada, K. *J. Antibiot.* **1987**, *40*, 450. (b) Yasuzawa, T.; Shirahata, C.; Sano, H. *J. Antibiot.* **1987**, *40*, 455. (3) (a) Sano, S.; Ikai, K.; Katayama, K.; Takesako, K.; Nakamura,

T.; Obayashi, A.; Ezure, Y.; Enomoto, H. J. Antibiot. 1986, 39, 1685. (b) Sano, S.; Ikai, K.; Kuroda, H.; Nakamura, T.; Obayashi, A.; Ezure, Y.; Enomoto, H. J. Antibiot. 1986, 39, 1674. (c) Sano, S.; Ikai, K.; Yoshikawa, Y.; Nakamura, T.; Obayashi, A. *J. Antibiot.* **1987**, *40*, 512. (d) Sano, S.; Kuroda, H.; Ueno, M.; Yoshikawa, Y. Nakamura, T.; Obayashi, A. J. Antibiot. 1987, 40, 519.

(4) Jolad, S. D.; Hoffmann, J. J.; Torrance, S. J.; Wiedhopf, R. M.; Cole, J. R.; Arora, S. K.; Bates, R. B.; Gargiuro, R. L.; Krieke, G. R. J. Am. Chem. Soc. 1977, 99, 8040.

(5) (a) Review: Itokawa, H.; Takeya, K. *Heterocycles* **1993**, *35*, 1467. (b) Ikokawa, H.; Takeya, K.; Mori, N.; Sonobe, T.; Mihashi, S.; Hamanaka, T. *Chem. Pharm. Bull.* **1986**, *34*, 3762. (c) Itokawa, H.; Yakeya, K.; Mihara, K.; Mori, N.; Hamanaka, T.; Sonobe, T.; Iitaka, Y. *Chem. Pharm. Bull.* **1983**, *31*, 1424. (d) Itokawa, H.; Takeya, K.;

Y. Chem. Pharm. Bull. 1983, 31, 1424. (d) Itokawa, H.; Takeya, K.;
Mori, N.; Kidohoro, S.; Yamamoto, H. Planta Med. 1984, 51, 313.
(6) Boger, D. L.; Yohannes, D. J. Org. Chem. 1987, 52, 5283.
(7) (a) Siuda, J. F. J. Org. Chem. 1975, 40, 3611. (b) Banerjee, N.;
Ressler, C. J. Org. Chem. 1976, 41, 3056. (c) Kawai, M.; Chorev, M.;
Marin-Rose, J.; Goodman, M. J. Med. Chem. 1980, 23, 420. (d) Cohen,
T.; Dietz, A. G., Jr.; Miser, J. R. J. Org. Chem. 1977, 42, 2057. (e) Konda, M.; Shioiri, T.; Yamada, S.-i. *Chem. Pharm. Bull.* **1975**, *42*, 2037. (e)
Konda, M.; Shioiri, T.; Yamada, S.-i. *Chem. Pharm. Bull.* **1975**, *23*, 1063. (f)
Bretshneider, H.; Hohenlohe-Oehringen, K.; Kaiser, A.; Wolcke, U. *Helv. Chim. Acta* **1973**, *56*, 2857. (8)
Jung, M. E.; Lazarova, T. I. *J. Org. Chem.* **1997**, *62*, 1553.

ester (8) from L-tyrosine (5) in six steps (34% overall yield). Boger's approach utilized an acid-promoted oxidative rearrangement of a benzylic hydroperoxide to obtain the target compound (Scheme 1).

Recently Jung and Lazarova reported a five-step synthesis of 4-O-benzyl-N-Boc-L-Dopa (12) from L-tyrosine (5) via sequential Reimer-Tiemann and Dakin reactions (Scheme 2).8 However, the Reimer-Tiemann formylation of N-Boc-tyrosine (9) required rather vigorous reaction conditions (7.5 equiv of NaOH, CHCl₃, reflux, 4 h) for a chiral amino acid,⁹ and gave a low yield of 10 (33% isolated yield, 64% yield based on 31% recovered stating material).

Although the Baeyer-Villiger oxidation of N-protected-3-acetyl-L-tyrosine esters is reported in poor yields $(\sim 30\%)$,¹⁰ the similar oxidation of 3-acetyl-L-tyrosine hydrochloride (13) proceeds in 75% yield to give undifferentiated L-Dopa.^{7f} We reasoned that it may be possible to achieve a higher yield of the desired phenol by optimizing reaction conditions for the Baeyer-Villiger oxidation of N-protected 3-acetyl-L-tyrosine or its ester derivative. Herein, we report an efficient method for the synthesis of selectively O-protected derivatives of L-Dopa, which can be utilized as suitable intermediates in the total synthesis of natural products such as compounds 2-3e.11 This method is based on a modified Baeyer-Villiger oxidation of N-protected 3-acetyl-L-tyrosine esters, resulting in a selective phenol introduction.¹²

3-Acetyl-L-tyrosine (13) was prepared according to Boger's protocol in 77% yield.⁶ Amine protection with Boc anhydride (Boc₂O, NaHCO₃, dioxane-H₂O, 94%) provided 14. Fully protected 3-acetyl-L-tyrosine 15 was obtained by converting the free phenol and the acid to the corresponding benzyl ether and benzyl ester respectively, utilizing the conditions described by Boger⁶ to minimize possible racemization (2.5 equiv of PhCH₂Br, 2.5 equiv of K₂CO₃, 0.1 equiv of Et₄NI, DMF, 25 °C, 91%). The final transformation was a Baeyer-Villiger oxidation of the acetophenone to provide the required phenol. We investigated several different conditions and chose the m-CPBA oxidation at room temperature. Thus, the acetyl-L-tyrosine 15 was treated with 2 equiv of m-chloroperbenzoic acid in dichloromethane at room temperature for 7 days. Although the oxidation was sluggish it was very

^{(1) (}a) Tamai, S.; Kaneda, M.; Nakamura, S. J. Antibiot. 1982, 35, 1130. (b) Kaneda, M.; Tamai, S.; Nakamura, S.; Hirata, T.; Kushi, Y.; Suga, T. J. Antibiot. 1982, 35, 1137.

⁽⁹⁾ No optical rotation or chiral analysis data was reported for these compounds by Jung and Lazarova.⁸ Boger and Yohannes observed that benzylation of tyrosine phenol under vigorous reaction conditions (2 equiv of K₂CO₃, acetone, 60 °C) results in substantial racemization (15-20%).⁶ Selectively protected catechol derivatives of D,L-Dopa have been prepared from isovanillin, see: Wilcox, M. E.; Wyler, H.; Mabry, T. J.; Dreiding, A. S. Helv. Chim. Acta 1965, 48, 252

⁽¹⁰⁾ Boger, D. L.; Coleman, R. S. J. Org. Chem. 1986, 51, 5436. also see ref 7a-e.

^{(11) (}a) Schmidt, U.; Weller, D.; Holder, A.; Lieberknecht, A. *Tetrahedron Lett.* **1988**, *29*, 3227. (b) Boger, D. L.; Yohnnes, D.; Zhou, J.; Patane, M. A. J. Am. Chem. Soc. 1993, 115, 3420. (c) Boger, D. L.; J., Fatalie, M. A. J. Am. Chem. Soc. 1993, 113, 3420. (f) Boger, D. L.;
 Yohannes, D. J. Org. Chem. 1991, 56, 1763. (e) Boger, D. L.;
 Yohannes, D. J. Org. Chem. 1992, 57, 1319. (f) Boger, D. L.;
 Yohannes, D. J. Org. Chem. 1989, 54, 2498. (h) Boger, D. L.;
 Yohannes, D. J. Org. Chem. 1989, 54, 2498. (h) Boger, D. L.; D. J. Org. Chem. 1990. 55. 6000.

⁽¹²⁾ Selectively protected L-Dopa derivatives have been reported. These methods have relied on the monoprotection of the unsymmetrical catechol of Dopa derivatives, ^{7a-c,e-f} diazotization of 3-amino-L-tyrosine derivatives and subsequent copper(I)-promoted phenol introduction,7d or Baeyer-Villiger oxidation of 3-acetyl-L-tyrosine derivatives.¹⁰ All these methods provide the desired compounds in yield 30% or less.



clean, and the corresponding acetoxy derivative 16 was isolated in 81% yield after a simple aqueous workup and crystallization. These conditions were tested on 0.1 to 10 g scales, and the yields were very consistent. Similar results were obtained when peracetic acid was used as oxidizing reagent (83% yield).¹³ Heating the reaction in dichloromethane solvent did not help since more oxidant was required for complete conversion and the yield was lower, consistent with results reported by Boger.⁶ The acetyl group was cleaved by treatment of the reaction mixture with methanolic ammonia for 1 h before workup to afford the desired L-Dopa **17** in an overall yield of 74% for the two steps (Scheme 3). Thus, in a four-step procedure, the selectively monoprotected L-Dopa derivative, N-Boc-L-3-(3-hydroxy-4-benzoxyphenyl)alanine (17), was synthesized from L-tyrosine (5) in 49% overall yield.

To determine the optical purity of the (L)-3-hydroxytyrosine derivatives, compound **16** was converted to L-3-(3-hydroxy-4-benzoxyphenyl)alanine (**18**) hydrochloride salt (4 N HCl in dioxane), which was then coupled with (*R*)-(+)- α -methoxy- α -trifluoromethylphenylacetic acid and (*S*)-(-)- α -methoxy- α -trifluoromethylphenylacetic acid (EDC, DMAP, DMF, rt) (Scheme 4). The two Mosher's amide derivatives¹⁴ (**19a** and **19b**) were characterized by both proton and fluorine NMR spectra, which showed a single isomer for both **19a** and **19b** (ee > 95%).



^a Reagents: (a) AcCl/AlCl₃/PhNO₂/100 °C, 77%; (b) $Boc_2O/NaHCO_3/dioxane-H_2O$, 94%; (c) $BnBr/K_2CO_3/Et_4NI(cat.)/DMF/rt$, 91%; (d) mCPBA/CH₂Cl₂, rt, 81%; or CH₃CO₃H/CH₂Cl₂/rt, 83%; (e) NH₃MeOH, rt, 74% from **15**.

In conclusion, we have described here an efficient synthesis of the important selectively protected L-Dopa derivative **17** from L-tyrosine (**5**) via a modified Baeyer–Villiger oxidation. This compound can be used in the synthesis of natural products such as compounds 2-3e.

Experimental Section

N-(tert-Butyloxycarbonyl)-3-(3-acetyl-4-hydroxyphenyl)-L-alanine (14). 1-3-(3-Acetyl-4-hydroxyphenyl)alanine hydrochloride⁶ (**13**, 18 g, 50 mmol) was suspended in dioxane-water (1:1, 250 mL) and treated portionwise with sodium bicarbonate (12.6 g, 150 mmol). This solution was then treated with Boc₂O (1.2 equiv), and the mixture was vigorously stirred at roomtemperature overnight before it was concentrated in vacuo to about 100 mL. This residue was acidified with 2 N HCl to pH 2, and the product was extracted with ethyl acetate. The extract was washed with brine, dried over MgSO₄, and concentrated in vacuo. The crude product was dissolved in hot dichloromethane and treated with hexanes to precipitate a white solid (23 g, 94%): mp 110-112 °C; [α]²²_D +21.6° (c 2.2, MeOH); ¹H NMR (CDCl₃, 300 MHz): 1.33 (s, 0.4×9 H), 1.42 (s, 0.6×9 H), 2.60 (s, 3H), 3.02 (m, 2H), 4.35 (m, 0.4H), 4.61 (m, 0.6H), 5.09 (d, J = 7.8 Hz, 0.6H), 6.74 (d, J = 6.9 Hz, 0.4H), 6.92 (d, J = 8.1 Hz, 1H), 7.30 (d, J = 8.1 Hz, 1H), 7.55 (s, 1H), 8.42 (brs, 2H); CI-MS, m/e 234 (M + H – Boc). Anal. Calcd for C₁₆H₂₁NO₆: C, 59.43; H, 6.55; N, 4.33. Found: C, 59.21; H, 6.93; N, 4.24.

N-(tert-Butyloxycarbonyl)-3-(3-acetyl-4-benzyloxyphenyl)-L-alanine Benzyl Ester (15). A solution of N-(tertbutyloxycarbonyl)-3-(3-acetyl-4-hydroxyphenyl)-L-alanine (14, 13 g, 40 mmol) in dry DMF (150 mL) at room temperature was treated with potassium carbonate (14 g, 100 mmol), benzyl bromide (17.1 g, 100 mmol), and tetraethylammonium iodide (1.2 g, 5 mmol). The resulting reaction mixture was stirred at roomtemperature overnight and partitioned into ether and water. The aqueous phase was extracted with ether, and the combined ether extract was washed with 1 N HCl and brine, dried over MgSO₄, and concentrated in vacuo. The crude product was chromatographed on silica gel to give the product as a colorless oil (18.3 g, 91% yield), $[\alpha]^{2\bar{2}}_{D}$ –2.9° (c 1.4, MeOH); ¹H NMR (CDCl₃, 300 MHz): 1.42 (s, 9H), 2.57 (s, 3H), 3.00 (dd, J = 6.3, 13.8 Hz, 1H), 3.07 (dd, J = 5.1, 13.8 Hz, 1H), 4.58 (m, 1H), 5.02 (d, J = 8.1Hz, 1H), 5.12 (s, 2H), 5.13 (d, J = 5.7 Hz, 1H), 5.15 (d, J = 5.7

⁽¹³⁾ On one millimole scale using peracetic acid (32 wt % in acetic acid, 2 equiv, room temperature, 10 days).

⁽¹⁴⁾ a) Erickson, S. D.; Simon, J. A.; Still, W. C. *J. Org. Chem.* **1993**, *58*, 1305. (b) Tsuda, Y.; Hosoi, S.; Ishida, K.; Sangai, M. *Chem. Pharm. Bull.* **1994**, *42*, 204.

Scheme 4^a



^a Reagents: (a) 4 N HCl in dioxane, rt; (b) EDC/DMAP/DMF/Et₃N/rt.

Hz, 1H), 6.88 (d, J = 8.4 Hz, 1H), 7.12 (dd, J = 2.4, 8.4 Hz, 1H), 7.38 (m, 10H), 7.51 (d, J = 2.4 Hz, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ 28.0, 31.1, 36.9, 54.3, 66.8, 70.4, 79.5, 112.7, 127.2, 127.8, 128.0, 128.1, 128.2, 128.3, 128.4, 128.6, 130.9, 134.0, 134.8, 135.7, 154.7, 162.0, 171.1, 198.7; CI-MS *m/e* 404 (M + H - Boc). Anal. Calcd for C₃₀H₃₃NO₆: C, 71.55; H, 6.61; N, 2.78. Found: C, 71.17; H, 6.70; N, 2.76.

N-(tert-Butyloxycarbonyl)-3-(3-acetoxy-4-benzyloxyphenyl)-L-alanine Benzyl Ester (16). A solution of N-(tertbutyloxycarbonyl)-3-(3-acetyl-4-benzyloxyphenyl)-L-alanine benzyl ester (15, 1.01 g, 2 mmol) and mCPBA (57-84%, 1.25 g, ~2 equiv) in dichloromethane (20 mL) was stirred at room temperature for 7 days. The solution was diluted with ether (200 mL) and washed with saturated sodium thiosulfate, saturated sodium bicarbonate, and brine, dried over sodium bicarbonate, filtered, and concentrated in vacuo. The residue was crystallized from ether—hexanes to give the product as a white solid (841 mg, 81% yield), mp 88–90 °C, $[\alpha]^{22}{}_{\rm D}$ –5.6° (c2.2, MeOH); $^1{\rm H}$ NMR (CDCl₃, 300 MHz): 1.56 (s, 9H), 2.26 (s, 3H), 3.02 (m, 2H), 4.58 (m, 1H), 4.99 (d, J = 8.1 Hz, 1H), 5.04 (s, 2H), 5.09 (d, J = 12.6 Hz, 1H), 5.17 (d, J = 12.6 Hz, 1H), 6.78 (s, 1H), 6.84 (s, 2H), 7.31 (m, 10H); $^{13}\mathrm{C}$ NMR (CDCl₃, 75 MHz) δ 20.6, 28.2, 37.2, 54.3, 66.9, 70.5, 79.8, 113.7, 123.5, 126.9, 127.3, 127.7, 128.2 (brs), 128.3, 128.7, 135.0, 136.5, 139.8, 149.0, 154.8, 168.6, 171.3; CI-MS m/e 420 (M + H – Boc). Anal. Calcd for $C_{30}H_{33}NO_7$: C, 69.35; H, 6.40; N, 2.70. Found: C, 69.18; H, 6.45; N, 2.68.

N-(tert-Butyloxycarbonyl)-3-(3-hydroxy-4-benzyloxyphenyl)-L-alanine Benzyl Ester (17). A solution of N-(tertbutyloxycarbonyl)-3-(3-acetyl-4-benzoxyphenyl)-L-alanine benzyl ester (15, 1.01 g, 2 mmol) was dissolved in dichloromethane (20 mL) and treated with m-chloroperbenzoic acid (57%-84%, 1.25 g). This solution was stirred at room temperature for 7 days. A $\tilde{2}$ M solution of ammonia in methanol ($\hat{4}$ mL) was added, and the mixture was stirred for another 1 h. The resultant mixture was concentrated in vacuo, and the residue was diluted with ether (200 mL) and washed with saturated sodium thiosulfate, saturated sodium bicarbonate and brine, dried over sodium bicarbonate, filtered, and concentrated in vacuo. The crude product was purified on silica gel with 1:2 ethyl acetate-hexanes to give the title compound as a colorless syrup (706 mg, 74% yield), $[\alpha]^{22}_{D} - 4.0^{\circ}$ (*c* 1.0, MeOH); ¹H NMR (CDCl₃, 300 MHz): 1.41 (s, 9H), 2.96 (d, J = 5.7 Hz, 2H), 4.56 (m, 1H), 5.02 (s, 2H), 5.04 (brs, 1H), 5.09 (d, J = 12.0 Hz, 1H). 5.14 (d, J = 12.0 Hz, 1H), 5.84 (brs, 1H), 6.48 (d, J = 8.4 Hz, 1H), 6.68 (d, J = 1.5 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 7.32 (m, 10H); ¹³C NMR (CDCl₃, 75 MHz) & 28.2, 37.5, 54.5, 66.9, 71.0, 79.8, 112.0, 115.7, 120.6, 126.7, 127.6, 128.1, 128.2, 128.3, 128.5, 129.2, 135.0, 136.1, 144.7, 145.0, 154.9, 171.5; CI-MS m/e 288 (M + H - Boc). Anal. Calcd for C28H31NO6: C, 70.42; H, 6.54; N, 2.93. Found: C, 70.18; H, 6.73; N, 2.76.

3-(3-Acetoxy-4-benzyloxyphenyl)-L-alanine Benzyl Ester Hydrochloride (18). A solution of N-(tert-butyloxycarbonyl)-3-(3-acetoxy-4-benzyloxyphenyl)-L-alanine benzyl ester (16, 2.5 g, 4.8 mmol) in 20 mL of CH₂Cl₂ was treated with 4 M HCl in dioxane (10 mL) and stirred overnight at room temperature under nitrogen. After overnight, some solid had formed, and the solution was triturated with 100 mL of ether. The suspension was stirred for 1 h and filtered to give a white solid, which was dried to yield compound 18 as a white solid, 1.86 g (85%); mp 194–6 °C; ¹H NMR (TMS/CD₃OD): 2.25 (s, 3H), 3.10 (dd, J =6.9, 13.8 Hz, 1H), 3.18 (dd, J = 6.3, 13.8 Hz, 1H), 4.31 (dd, J =6.3, 6.9 Hz, 1H), 5.09 (s, 2H), 5.23 (d, J = 10.8 Hz, 1H), 5.24 (d, J = 10.8 Hz, 1H), 6.92 (d, J = 2.1 Hz, 1H), 6.98 (d, J = 2.1, 8.4 Hz, 1H), 7.02 (d, J = 8.4 Hz, 1H), 7.35 (m, 10H); CIMS m/e 420 (M + 1). Anal. Calcd for C₂₅H₂₆ClNO₅: C, 65.86; H, 5.75; N, 3.07. Found: C, 65.82; H, 5.70; N, 3.00.

N-[(R)-(+)-α-Methoxy-α-trifluoromethylphenylacetyl]-3-(3-acetoxy-4-benzyloxyphenyl)-L-alanine Benzyl Ester (19a). A solution of 3-(3-acetoxy-4-benzyloxyphenyl)-L-alanine benzyl ester hydrochloride (18, 200 mg, 0.44 mmol), (R)-(+)-α-methoxy- α -trifluorophenylacetic acid (105 mg, 0.45 mmol), EDC (115 mg, 0.6 mmol), triethylamine (0.14 mL, 1.0 mmol), and a catalytic amount of DMAP in DMF (5 mL) was stirred under nitrogen for 3 days. The reaction mixture was partitioned between ether (100 mL) and 1 N HCl (30 mL). The organic layer was separated and washed with brine (2 \times 50 mL), dried over magnesium sulfate, and concentrated in vacuo. The product was purified on silica gel ($R_f = 0.6$, 30% EtOAc in hexane) to give the title compound as a colorless oil. ¹H NMR (TMS/CDCl₃): 2.26 (s, 3H), 3.08 (dd, J = 8.4, 14.0 Hz, 1H), 3.13 (dd, J =5.4, 14.0 Hz, 1H), 3.20 (d, $J_{\rm H,F} = 1.2$ Hz, 3H), 4.70 (brs, 1H), 4.90 (m, 1H), 5.05 (s, 2H), 5.10 (d, J = 12.0 Hz, 1H), 5.19 (d, J= 12.0 Hz, 1H), 6.80 (s, 1H), 6.84 (s, 2H), 7.36 (m, 14H), 7.48 (m, 1H); ¹⁹F NMR (CFCl₃/CDCl₃): -69.8; CIMS *m/e* 636 (M + 1); HRMS (FAB) calcd for C₃₅H₃₂F₃NO₇ m/e 636.2209, found 636.2230.

N-[(*S*)-(-)-α-Methoxy-α-trifluoromethylphenylacetyl]-3-(3-acetoxy-4-benzyloxyphenyl)-L-alanine Benzyl Ester (19b). The product was obtained by coupling of **18** with (S)-(-)-αmethoxy-α-trifluorophenylacetic acid as described above. Colorless oil; ¹H NMR (TMS/CDCl₃): 2.25 (s, 3H), 3.01 (m, 2H), 3.40 (d, J = 1.8 Hz, 3H), 4.99 (m, 1H), 5.01 (s, 2H), 5.11 (d, J = 12.0 Hz, 1H), 5.20 (d, J = 12.0 Hz, 1H), 6.53 (m, 2H), 6.66 (d, J = 9.0 Hz, 1H), 7.02 (m, 1H), 7.37 (m, 15H). ¹⁹F NMR (CFCl₃/CDCl₃): -69.1; CIMS *mle* 636 (M + 1); HRMS (FAB) calcd for C₃₅H₃₂F₃-NO₇ *mle* 636.2209, found 636.2229.

JO9913661